



This is the author's version of a work that was accepted for publication in the following source:

Halupka, K. J., M. N. Shivdasani, S. L. Cloherty, D. B. Grayden, Y. T. Wong, A. N. Burkitt, and H. Meffin. 2017. Prediction of cortical responses to simultaneous electrical stimulation of the retina. *Journal of Neural Engineering*. **14**(1): 016006.

**Notice:** Changes introduced as a result of publishing processes such as copy-editing and formatting may not be reflected in this document. For a definitive version of this work, please refer to the published source.

The final publication is available at:

<https://doi.org/10.1088/1741-2560/14/1/016006>

Copyright of this article belongs to: © 2016 IOP Publishing Ltd

Predicting responses to electrical retinal stimulation

# Prediction of Cortical Responses to Simultaneous Electrical Stimulation of the Retina

Abbreviated title: Predicting responses to electrical retinal stimulation

Kerry Halupka<sup>1,2,3</sup>, Mohit N. Shivdasani<sup>3,4</sup>, Shaun L. Cloherty<sup>5,6,1</sup>, David B. Grayden<sup>1,3,7</sup>, Yan T. Wong<sup>1</sup>, Anthony N. Burkitt<sup>1,3,7</sup>, Hamish Meffin<sup>5,6</sup>

<sup>1</sup>NeuroEngineering Laboratory, Dept. of Electrical & Electronic Engineering, The University of Melbourne, VIC 3010, Australia.

<sup>2</sup>National ICT Australia, Victoria Research Lab, The University of Melbourne, VIC 3010

<sup>3</sup>Bionics Institute, 384-388 Albert St, East Melbourne, VIC 3002, Australia.

<sup>4</sup>Dept. of Medical Bionics, The University of Melbourne, VIC 3010, Australia.

<sup>5</sup>National Vision Research Institute, Australian College of Optometry, Carlton, VIC 3053, Australia.

<sup>6</sup> Australian Research Council Centre of Excellence for Integrative Brain Function, Department of Optometry and Vision Sciences, The University of Melbourne, Melbourne, VIC 3010, Australia.

<sup>7</sup>Centre for Neural Engineering, The University of Melbourne, VIC 3010, Australia.

Correspondence should be addressed to:

Kerry Halupka

Tel: +61 437 911 079

E-mail: Kerry.halupka@gmail.com

Address:

NeuroEngineering Laboratory

Dept. of Electrical & Electronic Engineering

The University of Melbourne

VIC 3010

Australia.

Page Count 43

Figures 10

Tables 0

Multimedia/3D models 0

## Word counts

Abstract 263

Introduction 602

Discussion/Conclusion 2106

**Total (Abstract, Intro, Methods, Results, Discussion, Conclusion, Acknowledgements)**

**7400**

Predicting responses to electrical retinal stimulation

1

## 2 **Abstract**

### 3 *Objective*

4 Simultaneous electrical stimulation of multiple electrodes has shown promise in  
5 diversifying the responses that can be elicited by retinal prostheses compared to  
6 interleaved single electrode stimulation. However, the effects of interactions between  
7 electrodes are not well understood and clinical trials with simultaneous stimulation have  
8 produced inconsistent results. We investigated the effects of multiple electrode  
9 stimulation of the retina by developing a model of cortical responses to retinal  
10 stimulation.

### 11 *Approach*

12 Electrical stimuli consisting of temporally sparse, biphasic current pulses, with  
13 amplitudes sampled from a bi-dimensional Gaussian distribution, were simultaneously  
14 delivered to the retina across a 42-channel electrode array implanted in the  
15 suprachoroidal space of anesthetized cats. Visual cortex activity was recorded using  
16 penetrating microelectrode arrays. These data were used to identify a linear-nonlinear  
17 model of cortical responses to retinal stimulation. The ability of the model to generalize  
18 was tested by predicting responses to non-white patterned stimuli.

### 19 *Main results*

20 The model accurately predicted two cortical activity measures: multi-unit neural  
21 responses and evoked potential responses to white noise stimuli. The model also  
22 provides information about electrical receptive fields, including the relative effects of

Predicting responses to electrical retinal stimulation

1  
2  
3 1 each stimulating electrode on every recording site.  
4  
5

6 2 *Significance*  
7  
8

9 3 We have demonstrated a simple model that accurately describes cortical responses to  
10 4 simultaneous stimulation of a suprachoroidal retinal prosthesis. Overall, our results  
11 5 demonstrate that cortical responses to simultaneous multi-electrode stimulation of the  
12 6 retina are repeatable and predictable, and that interactions between electrodes during  
13 7 simultaneous stimulation are predominantly linear. The model shows promise for  
14 8 determining optimal stimulation paradigms for exploiting interactions between electrodes  
15 9 to shape neural activity, thereby improving outcomes for patients with retinal prostheses.  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25

26 10  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

## 1 Introduction

2 Retinal prostheses produce phosphenes by electrically stimulating surviving retinal  
3 neurons in patients with severe photoreceptor degeneration. Clinical trials have shown  
4 that retinal prostheses elicit useful perceptions, resulting in improvements in  
5 spatio-motor tasks (Barry et al., 2012; Kotecha et al., 2014; Stingl et al., 2015) and  
6 reading (da Cruz et al., 2013). However, spatial resolution with present devices is  
7 severely limited with the highest reported visual acuity achieved of 20/546 (Stingl et al.,  
8 2013).

9 One strategy that shows promise to improve resolution is simultaneous  
10 stimulation of multiple electrodes. Interactions that occur when combinations of  
11 electrodes are stimulated simultaneously are capable of increasing the repertoire of  
12 visual percepts that can be elicited compared to conventional single-electrode  
13 stimulation. Simultaneous stimulation can activate groups of cells between electrodes  
14 (Dumm et al., 2014), reduce current spread by using one or more local return electrodes  
15 (Cicione et al., 2012; Habib et al., 2013; Matteucci et al., 2013), and elicit responses at  
16 the level of a single cell *in vitro* (Jepson et al., 2014). Clinically, studies have mainly  
17 aimed to elucidate perceptual differences between sequential and simultaneous  
18 stimulation (Humayun et al., 1999; Rizzo et al., 2003; Auner et al., 2008; Horsager et al.,  
19 2010; Wilke et al., 2011). Although these studies were limited to stimulation of electrodes  
20 in simple geometric patterns such as lines and squares with identical biphasic pulses on  
21 all electrodes, inconsistencies were reported regarding percept predictability and  
22 reproducibility.

23 It is clear that simultaneous stimulation is a promising strategy for eliciting useful

Predicting responses to electrical retinal stimulation

1  
2  
3 1 percepts. However, the crucial next step in either reducing or harnessing electrode  
4  
5 2 interactions is to investigate the predictability and reproducibility of responses to  
6  
7 3 simultaneous stimulation, and gain a better understanding of how electrode interactions  
8  
9 4 affect neural responses.  
10  
11

12  
13 5 Here, we combine simultaneous stimulation of 42 suprachoroidal electrodes with  
14  
15 6 multi-site recordings in the cat visual cortex, to demonstrate that electrode interactions at  
16  
17 7 the level of the cortex are predominantly linear. Additionally, we show that a simple  
18  
19 8 model comprised of a linear filter followed by a static nonlinearity (hereafter,  
20  
21 9 linear-nonlinear model) can reliably predict neural spiking responses to several different  
22  
23 10 simultaneous stimulation paradigms. We also demonstrate the ability of the same model  
24  
25 11 to predict responses based on an alternative metric of cortical activity, namely, the power  
26  
27 12 in the evoked responses. The predictive success of our model shows promise for  
28  
29 13 efficiently determining optimal stimulation paradigms for shaping neural activity with a  
30  
31 14 retinal prosthesis.  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

## 1 **Methods**

2 Experiments were performed in eight normally-sighted, adult cats. All procedures were  
3 approved by the Bionics Institute Animal Research Ethics Committee (Projects  
4 12/255AB and 14/304AB).

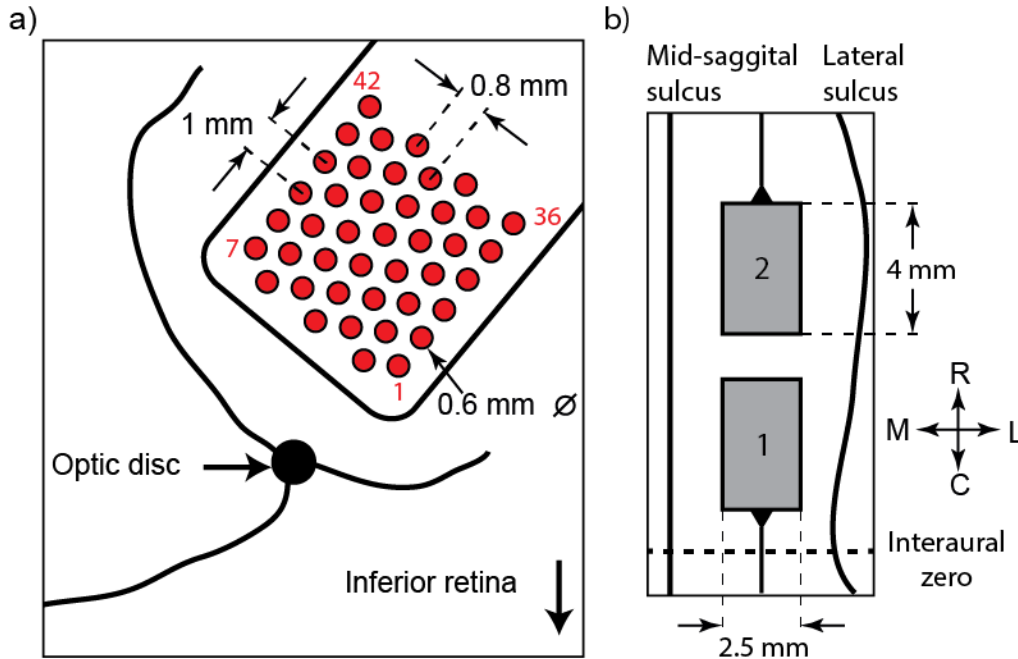
### 5 *Anesthesia and Surgery*

6 Cats were anesthetized with an initial dose of ketamine (intramuscular, 20 mg/kg) and  
7 xylazil (subcutaneous, 2 mg/kg). Anesthesia was maintained with an intravenous  
8 infusion of sodium pentobarbitone (60mg/kg/hr) and Hartmanns solution (sodium lactate,  
9 2.5 mL/kg/hour). Dexamethasone (intramuscular, 0.1 mg/kg) and Clavulox  
10 (subcutaneous, 10 mg/kg) injections were given daily to minimize brain swelling and  
11 infection. Vitals were continuously monitored throughout the experiment.

12 Details of the surgical implantation of the suprachoroidal array have been  
13 described previously (Shivdasani et al., 2012). Briefly, a lateral incision through the  
14 sclera was performed to expose the choroid. The electrode array was then inserted into  
15 the suprachoroidal space via a pocket created between the sclera and choroid, and  
16 sutured into place (Villalobos et al., 2012).

17 The suprachoroidal array (Figure 1(a)) consisted of a flexible medical grade  
18 silicone substrate with 7 rows x 3 columns of platinum electrodes used in six  
19 experiments, and 7 rows x 6 columns of electrodes used in the remaining two  
20 experiments. Electrodes were 600  $\mu\text{m}$  in diameter arranged hexagonally with 1 mm

## Predicting responses to electrical retinal stimulation



**Figure 1. a)** Schematic of stimulating array position and dimensions in relation to optic disc; electrode numbering for the four corner electrodes of the array are shown in red. **a)** Schematic of the recording array placements in the hemisphere of the visual cortex contralateral to the implanted eye. R: rostral, C: caudal, M: medial, L: lateral.

center-to-center spacing (Villalobos et al., 2012).

The visual cortex contralateral to the implanted eye was exposed and penetrating microelectrode arrays (either 6 x 10 or 6 x 6 channels, 1 mm length, 400  $\mu$ m spacing, Blackrock Micro., USA) were implanted Figure 1(b)). Evoked potentials (EPs) from the cortical surface in response to retinal stimulation were used to optimize the position of the implanted arrays (Cicione et al., 2012; Dumm et al., 2014). Neural signals were sampled at 30 kHz (Cerebus Neural Processing System, Blackrock Microsystems, Salt Lake City, UT).

### *White Noise Stimuli*

Predicting responses to electrical retinal stimulation

1  
2  
3 1 Temporally sparse electrical pulses delivered simultaneously across the stimulating  
4  
5 2 electrodes were used to characterize the system while responses were recorded in the  
6  
7 3 visual cortex. The stimulus amplitude for each electrode was sampled from a  
8  
9 4 bi-dimensional Gaussian distribution (hereafter, spatially white stimuli). To balance the  
10  
11 5 effect of each electrode on cortical activation, the standard deviation of each Gaussian  
12  
13 6 distribution was the activation threshold for that electrode.  
14  
15 7 Thresholds were acquired by stimulating each electrode individually with cathodic-first  
16  
17 8 biphasic current pulses using a 1 ms phase width, 25  $\mu$ s interphase gap, at a 1 Hz  
18  
19 9 presentation rate (the same pulse waveform was used for all electrical stimulation  
20  
21 10 paradigms), and 0-750  $\mu$ A stimulus amplitude, in 50  $\mu$ A steps. A spike-rate (or power)  
22  
23 11 versus stimulus amplitude input-output function for each stimulating electrode-recording  
24  
25 12 channel combination was generated to which a sigmoid curve was fitted. For each  
26  
27 13 sigmoid, the stimulus amplitude at 50% of the maximum response was defined as the  
28  
29 14 threshold for that recording channel (Shivdasani et al., 2012). The threshold for each  
30  
31 15 stimulating electrode was chosen to be the lowest threshold over all recording channels  
32  
33 16 for which a sigmoid could be fit. All electrodes across the array were then stimulated  
34  
35 17 simultaneously with charge-balanced waveforms using a mix of anodic and cathodic-first  
36  
37 18 polarities with current amplitudes determined by the stimulus matrix. There were 3600  
38  
39 19 white-noise stimulation patterns generated for each experiment; these were presented  
40  
41 20 eight times, with the average of the responses on each recording channel used for  
42  
43 21 analysis.

52  
53 22 To investigate variability in cortical responses to repeated white noise stimuli, 60  
54  
55 23 repetitions of 30 different white noise patterns were presented in a randomized order in  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

1  
2  
3 1 three of the experiments. The mean and standard deviation of the responses to each of  
4  
5 2 these patterns was calculated.

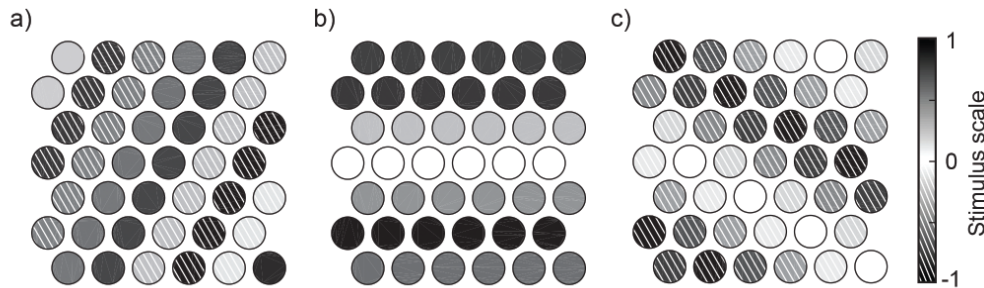
### 3 *Oriented Pattern Stimuli*

4 Stationary, oriented electrical stimulus patterns were presented to test the prediction  
5 ability of the model with non-white stimuli. These patterns were electrical representations  
6 of sinusoidal gratings at six possible orientations (0, 30, 60, 90, 120, and 150 degrees),  
7 five spatial frequencies (ranging over approximately 1-3 cycles per array length), and  
8 scaled by five stimulating currents (i.e., amplitudes of stimulating currents on each  
9 electrode were scaled by 175, 235, 295, 355, or 415  $\mu\text{A}$ ). Patterns consisted of three  
10 main types: (i) mixed-phase first stimuli (Figure 2(a)), where some electrodes on the  
11 array were stimulated with anodic-phase first biphasic pulses while others were  
12 stimulated with cathodic-phase first biphasic pulses; (ii) anodic-phase first stimuli (Figure  
13 2(b)), where all electrodes on the array were stimulated with anodic-phase first biphasic  
14 pulses; (iii) cathodic-phase first stimuli (Figure 2(c)), where all electrodes on the array  
15 were stimulated with cathodic-phase first biphasic pulses. During each experiment, 30  
16 different pattern stimuli were presented in a randomized order, with 10 repetitions of  
17 each pattern stimulus.

### 18 *Data Pre-Processing*

19 Two measures were used to estimate the cortical activity: power in the evoked potential  
20 (EP power) that occurred within approximately 20 ms of the stimulus pulse and multi-unit  
21 activity (MUA) in the same time frame.

## Predicting responses to electrical retinal stimulation



**Figure 2.** Example electrical stimulation patterns used as oriented electrical stimuli. The weighting of each electrode is represented by a grayscale, with black being the strongest weighting on the array (and thus the biggest current amplitude). White represents zero current amplitude. Electrodes with cathodic-phase first stimuli are hatched in white. **a)** Mixed-phase stimulus. **b)** Anodic-phase first only stimuli. **c)** Cathodic-phase first only stimuli.

To estimate the power in the evoked responses, offline multi-taper spectral analysis (Thomson, 1982; Mitra and Pesaran, 1999) was used to filter the raw data (400-1400 Hz, 2 tapers, taper length of 4 ms). The log power was then calculated from 3 ms after the end of the stimulus pulse (to exclude the stimulus artifact) until 20 ms after the pulse with the power in the same band in the 500 ms prior to each stimulus subtracted to exclude spontaneous activity.

For the MUA analyses, stimulus artefacts were first removed (Heffer and Fallon, 2008; Cicione et al., 2012; Shivdasani et al., 2012). Following band-pass filtering using a third-order Butterworth filter (300-5000 Hz), multiunit spikes were detected as threshold crossings using a threshold of 4x root-mean-square that was calculated within a 60 s moving time window. Typically, MUA occurred 3-20 ms post-stimulus; however, in some instances, another burst of spikes followed at approximately 30 ms post-stimulus. Only the early component of spiking was analyzed as this is considered to be a result of direct stimulation of retinal ganglion cells (RGCs) and indirect activation of the inner retina, but

Predicting responses to electrical retinal stimulation

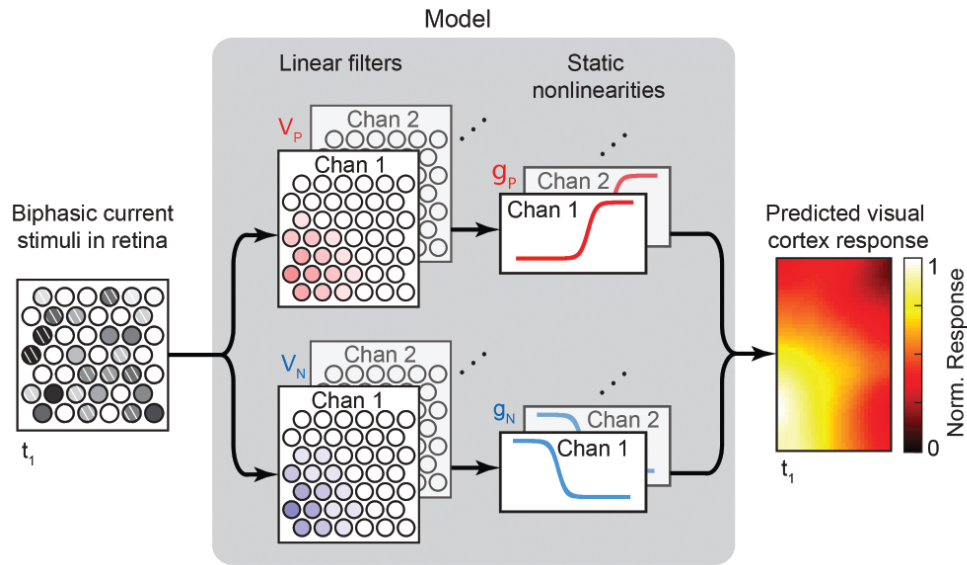
1 not activation of the photoreceptors (Boinagrov et al., 2014). The spontaneous spiking  
2 rate on each channel in the 500 ms prior to each stimulus was subtracted to account for  
3 variation in the stimulus-independent activity. For both measures, responses on each  
4 channel were normalized by the maximum response recorded on the array to any  
5 presented stimulus.

### 6 *Linear-Nonlinear (LN) Model*

7 We developed a model to explain activity in the visual cortex in response to  
8 simultaneous multi-electrode stimulation of the retina. Cortical responses to spatially  
9 white electrical stimulation of the retina were used to characterize the model. The model  
10 consisted of two parallel spatial linear filters that provided estimates of the spatial tuning  
11 properties of each cortical recording channel, followed by two parallel static  
12 nonlinearities that accounted for nonlinear characteristics of neurons such as response  
13 thresholds and saturation.

14 This model is similar to well-established Gaussian white noise models developed  
15 to describe light responses in the retina (Chichilnisky, 2001; Pillow et al., 2005; Schwartz  
16 and Rieke, 2011) and RGC responses to simultaneous stimulation of the retina  
17 (Maturana et al., 2016). However, several key adaptations have been made to the  
18 established model to accommodate the different stimulation method (i.e. electrical vs  
19 light stimulation). First, as opposite phase pulses produce a differential response  
20 (Jensen and Rizzo, 2006), the effects of both anodic-first and cathodic-first biphasic  
21 pulses were included separately in the model. Specifically, two different linear filters ( $V_P$   
22 for net anodic-first and  $V_N$  for net cathodic-first stimuli) and their associated static  
23 nonlinearities ( $g_P$  and  $g_N$ ) acted on the input ( $S$ ) separately (Figure 3).

## Predicting responses to electrical retinal stimulation



**Figure 3.** The linear non-linear cascade model of the pathway illustrated between the retina and visual cortex. Shown here is the input signal, arranged in the form of multiple electrode stimulation of the suprachoroidal array. The input is split by the model into a “positive” and a “negative” path, each representing the cortical response to net anodic first and net cathodic first stimuli respectively. After the linear and the nonlinear filters act on either side, the outputs are summed. The summed output is the predicted cortical response to a particular input stimulus. A total of 3600 white noise patterns were presented to the retina, here  $t_i$  refers to a single stimulus and its resulting predicted response.

Our model also accommodated the larger range of response amplitudes present in our data and adjusted to describe response amplitude, rather than spike probability. Additionally, we recorded cortical responses at multiple sites (up to 120) concurrently during stimulation and, as such, a LN model was fitted independently for each channel; thus, the index for each channel is omitted from the equations below for clarity.

An estimate of the activity (spike count or EP power) at each time  $t$  ( $t \in \{1 \dots T\}$ ),

Predicting responses to electrical retinal stimulation

1 at a given site in the cortex, is given by

$$\mathbf{R}_{\text{est}} = g_P(\mathbf{S}^T \mathbf{V}_P) + g_N(\mathbf{S}^T \mathbf{V}_N). \quad (1)$$

2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13 Each column of the stimulus matrix  $\mathbf{S}$  contains the stimulation amplitude applied  
14  
15 at time  $t$  with a row for each stimulating electrode  $j$  ( $j \in \{1 \dots M\}$ ), where  $T$  is the  
16  
17 number of stimuli that resulted in a response; the stimulus vector at time  $t$  is thus given  
18  
19  
20  
21 by

$$\mathbf{s}_t = \begin{pmatrix} s_{1,t} \\ \vdots \\ s_{M,t} \end{pmatrix}. \quad (2)$$

22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

8 The normalized stimulus vector is thus given by

$$\bar{\mathbf{s}}_t = \begin{pmatrix} s_{1,t}/\sigma_1 \\ \vdots \\ s_{M,t}/\sigma_M \end{pmatrix}. \quad (3)$$

9 where  $\sigma_M$  is the activation threshold of each stimulating electrode. Since more than one  
10 spike was sometimes recorded per stimulus, the spike triggered covariance is given by

$$\mathbf{C}_{\text{STC}} = \frac{1}{N} \sum_{t=1}^T r_{i,t} \bar{\mathbf{s}}_t \bar{\mathbf{s}}_t^T. \quad (4)$$

11 where  $r_{i,t}$  is the response on recording channel  $i$  ( $\{i \in 1 \dots C\}$ ), at time  $t$ , and  $N$  is the  
12 sum of responses on that channel.

13 The combined effect of all stimulating electrodes on a response in a given cortical  
14 channel could be either net anodic-first or net cathodic-first. Therefore, the effects of both  
15 anodic-first and cathodic-first biphasic pulses were included separately in the model.  
16 Responses due to a net anodic-first stimulation ( $\mathbf{R}_P$ ) were those where the projections of

Predicting responses to electrical retinal stimulation

1 the stimulus onto the principal eigenvector of  $\mathbf{C}_{\text{STC}}$  (denoted  $\mathbf{u}_{\text{STC}}$ ) were positive. Stimuli  
 2 for which this held true were denoted  $\bar{\mathbf{s}}_p$ , such that

$$\bar{\mathbf{s}}_p \cdot \mathbf{u}_{\text{STC}} > \mathbf{0}. \quad (5)$$

3 Responses to a net cathodic-first stimulation ( $\mathbf{R}_N$ ) were those for which the projections  
 4 onto the principal eigenvector were negative,

$$\bar{\mathbf{s}}_N \cdot \mathbf{u}_{\text{STC}} < \mathbf{0}, \quad (6)$$

5 where  $\bar{\mathbf{s}}_N$  were the corresponding cathodic-first stimuli.

6 The linear filter estimates  $\mathbf{V}_{P,\text{est}}$  and  $\mathbf{V}_{N,\text{est}}$  were then calculated as the spike  
 7 triggered average (Chichilnisky, 2001) of response subsets  $\mathbf{r}_p$  and  $\mathbf{r}_N$  and their  
 8 respective stimuli ( $\mathbf{s}_p$  and  $\mathbf{s}_N$ ),

$$\mathbf{V}_{P,\text{est}} = \frac{\mathbf{r}_p^T \bar{\mathbf{s}}_p}{N_p} \quad (7)$$

9 and

$$\mathbf{V}_{N,\text{est}} = \frac{\mathbf{r}_N^T \bar{\mathbf{s}}_N}{N_N}, \quad (8)$$

10 where  $N_p$  and  $N_N$  are the sum of response subsets  $\mathbf{r}_p$  and  $\mathbf{r}_N$ , respectively, such that  
 11  $N_p + N_N = N$ . Filters were then normalized by dividing by their L-2 norm. These  $\mathbf{V}_{P,\text{est}}$   
 12 and  $\mathbf{V}_{N,\text{est}}$  vectors were considered to be weightings for the effect of the stimulating  
 13 electrodes on a given cortical site.

14 The static nonlinearities  $g_p$  and  $g_N$  were approximated by sigmoids and  
 15 parameterized by their saturation amplitudes ( $y_p$  and  $y_N$ ), thresholds ( $a_p$  and  $a_N$ )  
 16 which are 50% of the saturation level, as well as the gain of the sigmoids ( $b_p$  and  $b_N$ ).  
 17 The equations for the static nonlinearities  $g_p$  and  $g_N$  are

Predicting responses to electrical retinal stimulation

$$g_P(r) = \frac{y_P}{1 + e^{(-b_P(x_P - a_P))}} \quad (9)$$

$$g_N(r) = y_N - \frac{y_N}{1 + e^{(-b_N(x_N - a_N))}}. \quad (10)$$

1 We used the distribution of the outputs of the linear filters,  $x_P = \bar{s}_P \mathbf{V}_{P,\text{est}}$  and  
 2  $x_N = \bar{s}_N \mathbf{V}_{N,\text{est}}$  (hereafter referred to as generator signals), and corresponding responses  
 3  $r_P$  and  $r_N$ , to provide an initial estimate for the parameters of non-linear functions  $g_P$   
 4 and  $g_N$  respectively. The threshold values  $a_P$  and  $a_N$  were estimated to be the mean  
 5 values of their respective generator signals, while  $y_P$  and  $y_N$  were estimated as that  
 6 maximum average response recorded on any channel (averaged over repeated trials).  
 7 We used a Levenberg-Marquardt non-linear least squares algorithm in MATLAB  
 8 (Mathworks, Inc. USA) to find the optimal non-linear parameters ( $a_P$ ,  $a_N$ ,  $b_P$ ,  $b_N$ ,  $y_P$   
 9 and  $y_N$ ) and confirm that the optimal linear filters were  $\mathbf{V}_{P,\text{est}}$  and  $\mathbf{V}_{N,\text{est}}$ . The outcome  
 10 of this process was a set of linear filters and static nonlinearities, each corresponding to  
 11 a different recording channel (shown for one recording channel in Figure 5).  
 12 To test which electrodes significantly affected a recording channel's response, we  
 13 randomly time-shifted the response vector 1000 times and repeated the above analysis  
 14 (Equations 1-8) for each new response subset. This generated a distribution for  $\mathbf{V}_{P,\text{est}}$   
 15 and  $\mathbf{V}_{N,\text{est}}$  to which the true vectors could be compared. Electrodes from the true  $\mathbf{V}_{P,\text{est}}$   
 16 and  $\mathbf{V}_{N,\text{est}}$  that were larger than the root mean square of the randomly-generated  
 17 distribution were considered significant. The spatial extent in the retina over which a  
 18 cortical site is influenced by is given by:

$$D_P = \frac{\sum_{j=1}^M v_j^P d_j}{v_j^P} \quad (11)$$

Predicting responses to electrical retinal stimulation

$$D_N = \frac{\sum_{j=1}^M v_j^N d_j}{v_j^N}, \quad (12)$$

1 where  $d_j$  is the distance of each electrode  $j$  to the center of mass of the significant  
 2 electrodes, and  $v_j^P$  and  $v_j^N$  are the weights given by  $V_{P,est}$  and  $V_{N,est}$ , respectively.

### 3 *Statistical Analysis*

4 To compare the sizes of  $V_{P,est}$  and  $V_{N,est}$ , we conducted a sign test to determine  
 5 whether the results of subtracting the filters for each channel were significantly different  
 6 to zero. False Discovery Rate correction (Benjamini and Hochberg, 1995) was used to  
 7 control for multiple comparisons to a level of  $p < 0.05$ . We used the same analysis to  
 8 compare the sizes of  $V_{P,est}$  and  $V_{N,est}$  between the two model types (MUA model and  
 9 EP power model).

10 To demonstrate the ability of the model to predict cortical responses to spatially white  
 11 electrical retinal stimulation, we performed a 6-fold cross validation analysis on all eight  
 12 experiments for models fitted to both MUA and evoked response power. For this, the  
 13 model was fitted using only 5/6 of the white noise data and used to predict the remaining  
 14 1/6 of the data. Only recording sites that exhibited a monotonic increase of activity with  
 15 current in response to single electrode stimulation of at least one stimulating electrode  
 16 were included in model fitting and analysis. Cross-correlation analyses were performed  
 17 to compare the predicted and recorded responses. The coefficient of determination and  
 18 slope of the line of best fit (where the predicted response was the independent variable)  
 19 were calculated for each of the cross-validation sets independently. Unless otherwise  
 20 stated, all figures with error bars show the mean and standard error of the mean.  
 21 Significant differences between the prediction abilities of the model fitted to MUA versus

Predicting responses to electrical retinal stimulation

1  
2  
3 1 that fitted to evoked response power were tested using a Wilcoxon Rank sum test ( $p <$   
4  
5  
6 2 0.05).

7  
8 3 In order to rule out any correlations due to chance, we estimated the number of  
9  
10 4 responses to white noise patterns in the trial set that could be accurately predicted by the  
11  
12 5 model. A bootstrap test was performed (10,000 samples) comparing the absolute  
13  
14 6 residual error of prediction for each white noise pattern in the trial set against a  
15  
16 7 distribution composed of the absolute residual error of recorded responses versus  
17  
18 8 time-shifted predictions, over all fitted channels. False Discovery Rate correction  
19  
20 9 (Benjamini and Hochberg, 1995) was used to control for multiple comparisons to a level  
21  
22 10 of  $p < 0.05$ .

23  
24  
25  
26  
27 11 We then tested the ability of a model trained with responses to white-noise stimuli to  
28  
29 12 predict responses to the oriented pattern stimuli. Models used in these predictions were  
30  
31 13 fitted to all of the responses to white-noise stimuli (rather than just 5/6 of the data). Here,  
32  
33 14 the coefficient of determination and slope of the line of best fit was calculated for each of  
34  
35 15 the five stimulation amplitudes separately for all eight experiments. Further bootstrap  
36  
37 16 analyses similar to those performed with the white noise data were also performed on  
38  
39 17 the pattern stimuli data to rule out any correlations that may have occurred due to  
40  
41 18 chance. The same analysis was performed to compare the predicted and measured  
42  
43 19 responses to single electrode stimulation, using the data obtained to calculate single  
44  
45 20 electrode thresholds. In addition to comparing the absolute residual error to the  
46  
47 21 bootstrapped distribution, the coefficient of determination between the predicted and  
48  
49 22 recorded responses to above-threshold single electrode stimulation was also compared  
50  
51 23 in the same manner.  
52  
53  
54  
55  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

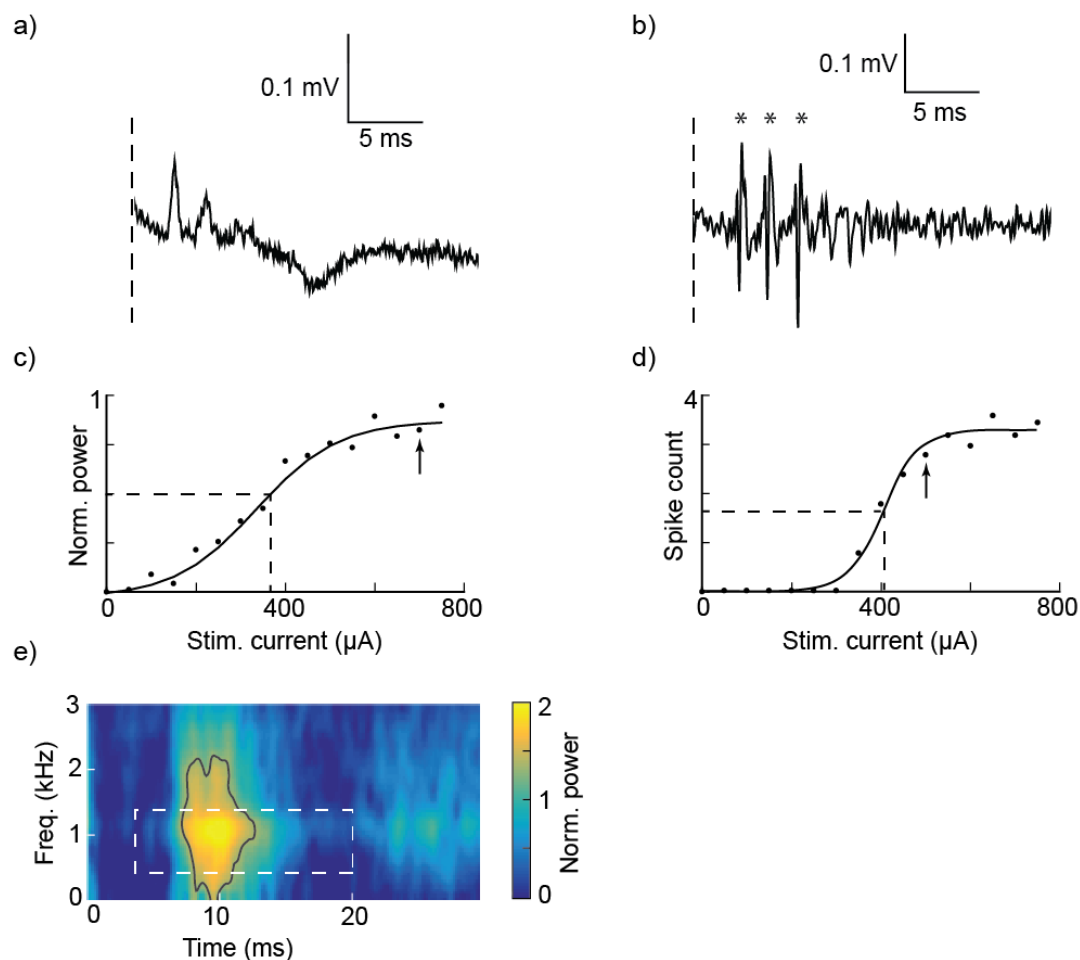
## 1 Results

2 Across eight experiments, visual cortex responses to multi-electrode stimulation of the  
3 retina were recorded and analyzed in the form of MUA and EP power on a total of 696  
4 recording channels.

### 5 *Cortical Responses to Multi-Electrode Suprachoroidal Stimulation*

6  
7 Figures 4(a) and 4(b) show raw recording traces from two recording sites following a  
8 stimulus pulse, indicating evoked responses, and the presence of spiking activity,  
9 respectively (to a white noise stimulus). Evoked responses typically consisted of two  
10 prominent peaks occurring a few milliseconds apart (Fig. 4(a)), sometimes followed by a  
11 third peak, resulting in a significant increase in power from the baseline response (Fig.  
12 4(e)). Both response measures exhibited a characteristic, monotonically increasing  
13 sigmoidal relationship with current amplitude (Figure 4(c,d)) (Cicione et al., 2012;  
14 Shivdasani et al., 2012; Dumm et al., 2014). However, significantly more channels  
15 exhibited this effect for EP power than MUA (MUA vs. EP power:  $66.6\% \pm 6.1$  vs.  $92.4\%$   
16  $\pm 3.2$ ,  $p = 0.0029$ ; Rank sum test;  $N=8$ ).

## Predicting responses to electrical retinal stimulation



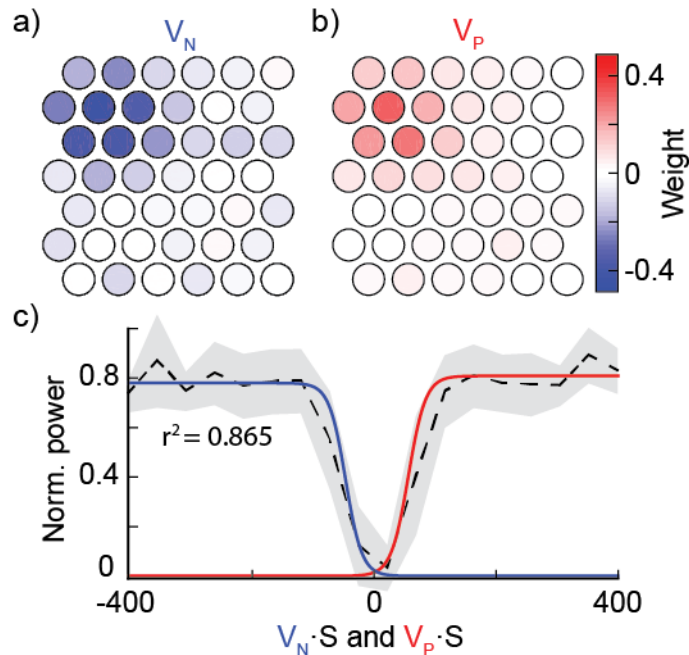
**Figure 4.** **a)** Raw response on one recording channel from a single electrode stimulus at 700  $\mu\text{A}$ , showing a multi-peaked evoked potential; dashed line shows the end of the stimulus artefact. **b)** Filtered trace on a different recording channel, showing spiking (indicated by asterisks) in response to single electrode stimulation. **c, d)** Transfer functions of the power in the multi-peaked evoked potential (c) and spike count (d) versus stimulation amplitude, with a sigmoidal line of best fit. Dashed lines show the thresholds of these particular stimulating electrodes at 50% of maximum response. The arrows show the average normalized power and spike count for responses shown in panels a and b respectively. **e)** Power spectrum of the multi-peaked response shown in panel a. The black outline indicates significant increase in power from the baseline response. White dashed box indicates the time and frequency window within which the EP power is calculated.

Predicting responses to electrical retinal stimulation

### 1 *Prediction of Cortical Responses*

2 Models for the response on each channel were fitted independently, and parametrized  
3 by two linear filters and two static nonlinearities (Figure 5). The linear filters derived  
4 analytically (Equations 2-8) and the optimized linear filters were compared, showing that  
5 the filters were largely identical before and after optimization, with a median  $r^2$  correlation  
6 coefficient of 0.93. The optimized linear filters and nonlinearity parameters were used in  
7 all further analyses. Figure 5 shows the components of the model for one recording  
8 channel. As shown by the linear filters, which can be considered to be the electrical  
9 receptive fields (ERFs) for each cortical channel, this particular recording site was  
10 maximally activated by pulses on the top left edge of the stimulating array. The static  
11 nonlinearities are functions of the action of the linear filters on the stimulus vector. For  
12 this recording channel, the model prediction of the responses (Figure 5(c)) was in close  
13 agreement with the recorded responses shown in grey (mean  $\pm$  SEM of the binned  
14 responses) (coefficient of determination  $r^2=0.865$ ).

Predicting responses to electrical retinal stimulation

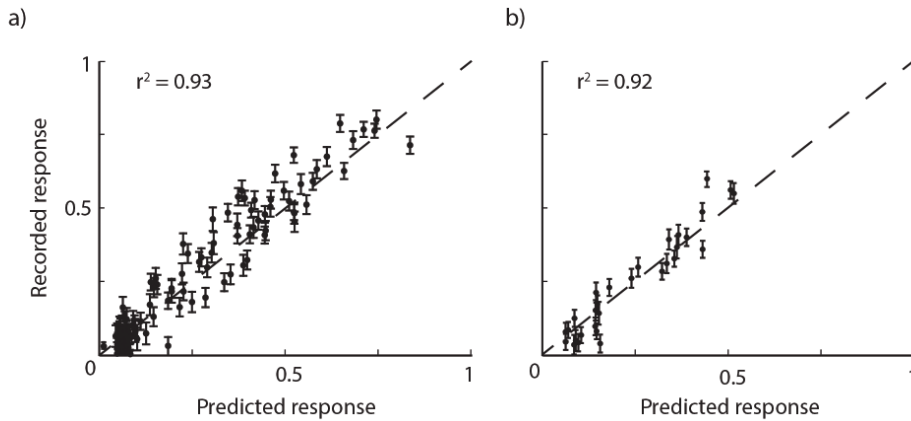


**Figure 5.** Model components for a single recording channel, as shown schematically in Figure 3, fitted with EP power in response to 5/6<sup>th</sup> of the white noise stimuli in one experiment. **a)** Spatial linear filters  $V_N$  and **b)**  $V_P$ , where each circle represents a stimulating electrode on the retinal array, and color refers to the strength of the linear filter. **c)** Static nonlinearities  $g_N$  (blue line) and  $g_P$  (red line). The y axis is normalized to the maximum average EP power recorded on any channel. Dashed black line shows the average recorded response to each of the binned generator signal levels (mean  $\pm$  SEM, shown in grey) ( $r^2=0.865$ ).

### Validating Model Fit

For the averaged responses to stimuli to be a reasonable metric to fit the model, the stimulus response variability must be minimal. Figure 6(a) shows the measured EP power (mean  $\pm$  SE) for all recording channels in response to 60 repetitions of a white noise stimulus pattern in one experiment. Only minor variation in responses was observed, and a clear and significant correlation was seen with the responses predicted

## Predicting responses to electrical retinal stimulation

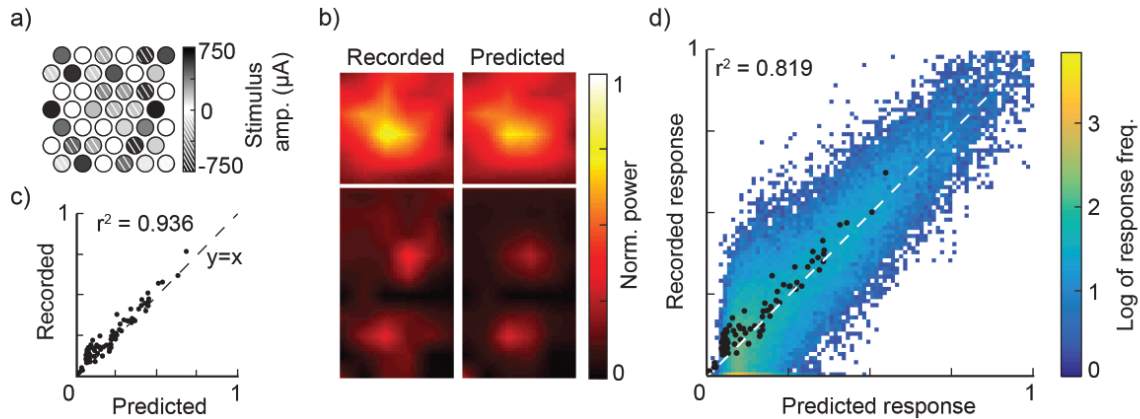


**Figure 6.** Mean and standard error of recorded responses ( $n=60$  repetitions of each stimulus) plotted against the responses predicted by the model. Response measure is power in the multi-peaked EP. a) Responses on 88 cortical channels in one experiment to 60 repetitions of a single white noise stimulus. b) Responses on a single recording channel in the same experiment as in panel a, to 60 repetitions of 30 separate white noise stimuli.

by the model ( $r^2 = 0.93$ ,  $p < 0.0001$ , FDR corrected permutation test, 10 million permutations). The same metric, but for a single cortical channel in response to 30 different white noise stimuli each repeated 60 times, is shown in Figure 6(b). Again, only minor variation in response was observed, with a clear and significant correlation ( $r^2 = 0.92$ ,  $p < 0.0001$ , FDR corrected permutation test, 10 million permutations).

The stimuli used to fit the model comprised 3600 different white noise patterns, averaged over eight repeats. For one such white noise stimulus (Figure 7(a)), a comparison of the recorded and predicted responses (Figure 7(b)) indicates that the model prediction of the response was very similar to the measured response, which was borne out by a coefficient of determination of  $r^2=0.936$  (Figure 7(c)), with a strong overall correlation observed for this experiment ( $r^2=0.819$ , Figure 7(d)).

## Predicting responses to electrical retinal stimulation



**Figure 7.** **a)** Example of a white-noise stimulus in one experiment with positive currents indicating anodic-first pulses and negative currents indicating cathodic-first pulses. **b)** Recorded and predicted EP power in the visual cortex across two recording arrays in response to the white noise stimulus shown in panel a (averaged over 8 repetitions). **c)** Correlation between the recorded response to the stimulus and the response predicted by the model. Each point represents the response measured on a single recording channel. Dashed line is  $y=x$ . **d)** Aggregated scatter plot for all 600 white-noise stimuli that were not used to fit the model for this experiment, with the log frequency of responses represented on the color scale. Black dots show the responses presented in panel c.

Similar correlations were found for all six cross-validated models for all eight experiments, with  $0.55 < r^2 < 0.89$  ( $n=48$ ). Models fitted to MUA displayed significantly higher correlation coefficients (MUA vs. EP power:  $r^2 = 0.77 \pm 0.01$  vs.  $0.69 \pm 0.02$ ,  $p=1.22 \times 10^{-4}$ ; Rank sum test) than those fitted to EP power. The average slope of the line of best fit for the models fitted to power was significantly closer to 1 (MUA vs. EP Power:  $0.88 \pm 0.018$  vs.  $0.92 \pm 0.13$ ,  $p=2.13 \times 10^{-4}$ ; Rank sum test) than those fitted to MUA.

Both types of models were able to predict the responses to at least 75% of the test set significantly better than chance (via bootstrap analysis), with the MUA model

Predicting responses to electrical retinal stimulation

1 predicting significantly more than the EP power model (MUA vs. EP Power:  $95.77\% \pm$   
2  $0.57\%$  vs.  $92.70\% \pm 0.90\%$ ,  $p=4.08 \times 10^{-4}$ ; Rank sum test).

### 3 4 *Electrical Receptive Fields Characterized by Linear Filter component*

5 The linear filters of the model provided an estimate of the ERFs of each recording  
6 channel for simultaneous retinal stimulation. Figure 8(a) shows the ERF for stimuli with a  
7 net cathodic-first effect, for the most rostro-medial recording site; this channel was most  
8 responsive to stimulation of the superior-nasal retinal electrodes. The interpolated color  
9 maps depicting the linear filters at all cortical sites ( $n=96$ ) from this experiment (Figure  
10 8(b)) display a clear preference to stimulation of superior-nasal retinal electrodes for  
11 more rostral recording sites, while caudal recording sites showed a preference for  
12 stimulation of electrodes located towards the inferior retina. The gradual movement of  
13 the ERFs between these two extremes indicates topographic mapping, and is likely to be  
14 retinotopic. Using these ERFs in combination with the static nonlinearities, it is possible  
15 to link regions of high activity to stimulus features. For example, the high power in the  
16 rostral (rostral direction shown in Figure 1(b)) sites in Figure 7(b) can be attributed to  
17 high stimulation amplitude on superior-nasal retinal electrodes. Figure 8(c) more clearly  
18 illustrates the rostro-caudal movement of the linear filters, where contours represent the  
19 significant linear filter weights belonging to the most rostro-medial (green) and  
20 caudo-medial (purple) recording sites (circled in Figure 8(b)). The recording sites in this  
21 example were separated by a distance of approximately 7 mm in the cortex, which  
22 corresponds to approximately  $14\text{-}23^\circ$  of visual angle using a cortical magnification factor  
23 of  $0.3\text{-}0.5$  mm/degree (Tusa et al., 1978). The peak weightings of the linear filters for  
24 these channels were separated by about 4 mm in retinal space, which subtends a visual

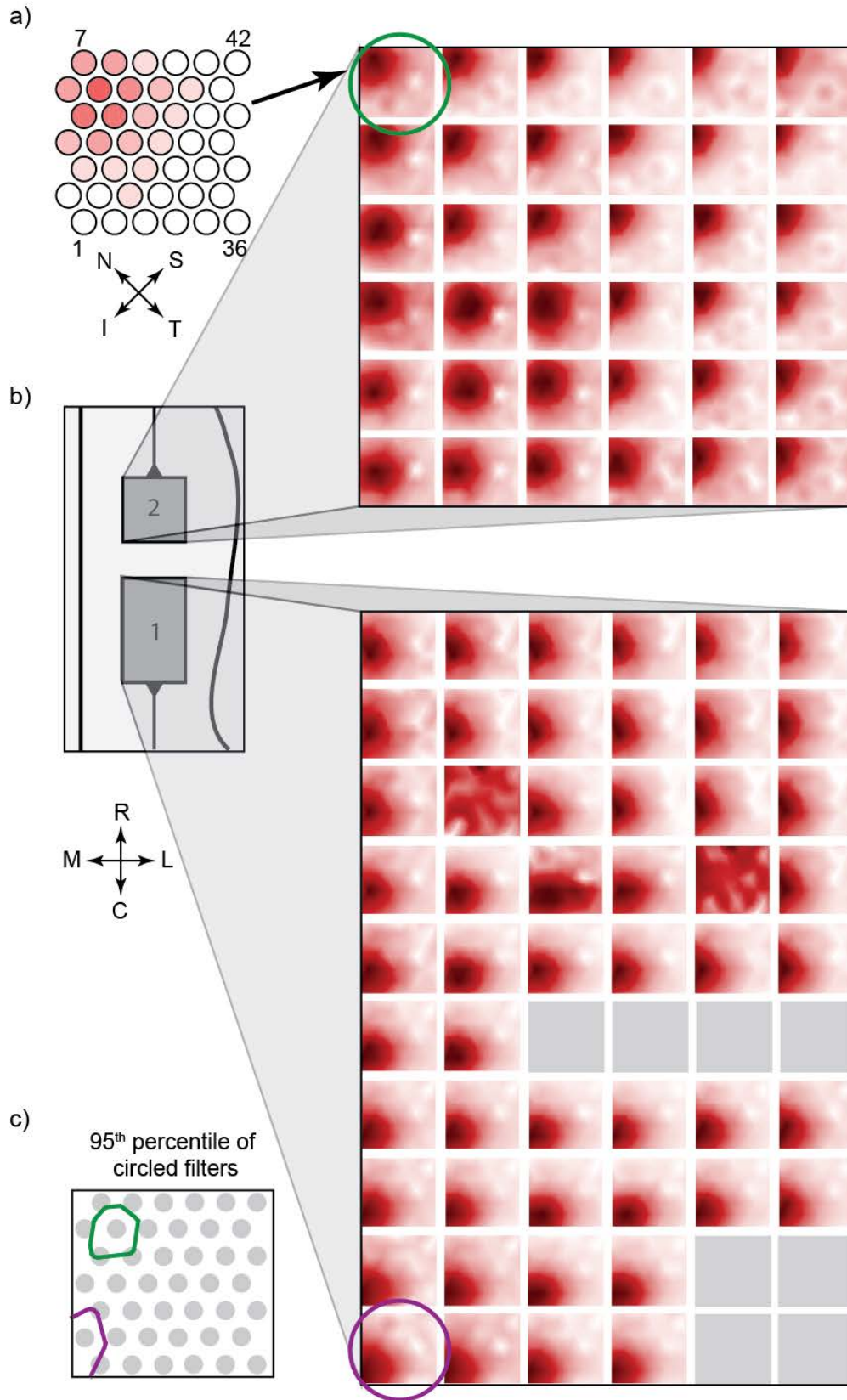
Predicting responses to electrical retinal stimulation

1  
2  
3 1 angle of approximately  $18^\circ$ , and thus corresponds well with the recording channel  
4  
5 2 separation.

6  
7  
8 3 While significant differences did occur between the size of ERFs for stimuli with a net  
9  
10 4 cathodic-first effect and those with a net anodic-first effect on a channel by channel basis  
11  
12 5 when using the EP power based model (Sign test, 0.05 level of significance), these  
13  
14 6 differences were not consistent across experiments. Two out of eight experiments had  
15  
16 7 larger  $V_N$  filters than  $V_P$  filters; however, one experiment had significantly larger  $V_P$   
17  
18 8 filters than  $V_N$  filters. No differences occurred between MUA-based model ERF's.  
19  
20 9 Similar inconsistent differences occurred when comparing the ERF size between the two  
21  
22 10 model types (EP power based models: two experiments had larger  $V_N$  filters, three had  
23  
24 11 larger  $V_P$  filters; MUA-based models: three experiments had larger  $V_N$  filters, three had  
25  
26 12 larger  $V_P$  filters).  
27  
28  
29  
30  
31

32 13  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation



1

2

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

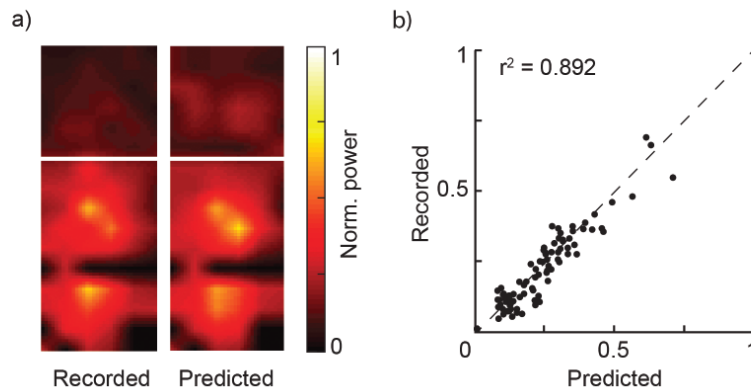
Predicting responses to electrical retinal stimulation

**Figure 8.** Linear filter maps for each recording site in one experiment. **a)** Linear filter of the most rostro-caudally located cortical site (marked with a black circle in panel b) prior to interpolation, as described in Figure 5. Only filters corresponding to anodic first stimulation (i.e.  $V_p$ ) are shown. Electrode numbers correspond to those indicated in Figure 1(a). S: superior retina, I: inferior retina, N: nasal retina, T: temporal retina. **b)** Interpolated color maps representing optimized linear filters for anodic first stimulation pulses ( $V_p$ ) corresponding to each of the 96 recording sites. Schematic of cortical recording areas corresponding to each of the linear filter maps is also shown. Red indicates a strong effect of a particular stimulating electrode on the activity recorded on a given cortical site. Blank sites are those that did not record a monotonically increasing response to single electrode stimulation and were thus excluded from analyses. R: rostral, C: caudal, M: medial, L: lateral. **c)** Schematic of stimulating array showing significant areas of the linear filter maps for two cortical recording sites marked with green and purple circles (corresponding to rostro-medial and caudo-medial recording sites respectively) in panel b. The shift in the electrodes that have the strongest contributions to a channels activity with cortical channel position shows that the retinotopic organization of the visual cortex is preserved with responses to simultaneous electrical stimulation of the retina.

### *Predicting Responses to Electrical Pattern Stimuli*

To examine whether a model fitted using responses to white noise stimuli could predict responses to non-white stimuli, measured responses to oriented pattern stimuli and single electrode stimuli were compared to responses predicted by the model. Three types of patterned stimuli were used: mixed-phase, anodic-first and cathodic-first. Figure 9(a) shows the comparison between the recorded and predicted responses for the stimulus shown in Figure 2(a), where the maximum current amplitude delivered was  $295\mu\text{A}$ . The responses predicted by the model corresponded well to the measured

## Predicting responses to electrical retinal stimulation

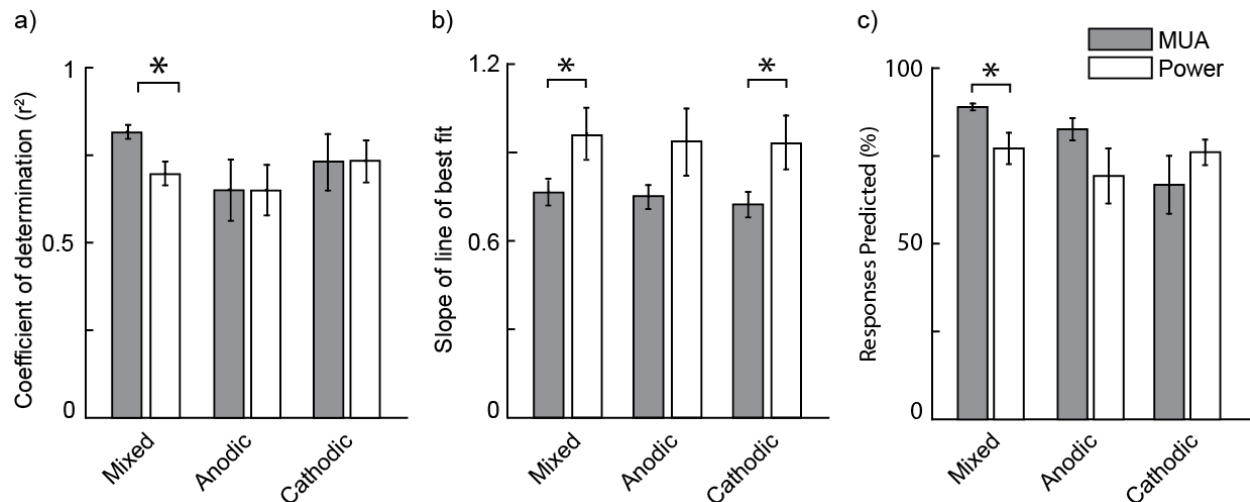


**Figure 9.** Recorded response and model-predicted response to the oriented pattern stimulus shown in Figure 2(a) (mixed phase first stimulus). Response is measured by the power in the EP, and shown over two recording arrays arranged as in Figure 8. The color map is normalized to the maximum recorded power. **a)** Recorded response (left), and predicted response (right). **b)** Correlation between the recorded and predicted responses (n=88 channels).

responses (Figure 9(b),  $r^2 = 0.89$ ) for this stimulus.

To assess whether this effect extended to the entire set of oriented pattern stimuli, we calculated a coefficient of determination for each of the three pattern types for all experiments (Figure 10(a)). Coefficients corresponding to MUA models were significantly higher than those for EP power models for mixed-phase stimuli (MUA vs. EP power:  $r^2 = 0.81 \pm 0.02$  vs.  $0.69 \pm 0.03$ ,  $p = 0.016$ ; Rank sum test). There was no difference between the MUA model and EP power model for the other two stimulus types (MUA vs. EP power with anodic-first pulses:  $r^2 = 0.65 \pm 0.07$  vs.  $0.65 \pm 0.05$ ,  $p = 0.558$ . MUA vs. EP power with cathodic-first pulses:  $0.73 \pm 0.06$  vs.  $0.73 \pm 0.04$ ,  $p = 0.457$ ). However, for the mixed-phase and cathodic-first stimulus types, the slopes of the lines of best fit for the models fitted to MUA were significantly lower than those fitted to EP power (Figure 10(b), MUA vs. EP power with mixed-phase pulses:  $0.76 \pm 0.05$  vs.  $0.96 \pm 0.09$ ,  $p = 0.02$ ).

## Predicting responses to electrical retinal stimulation



**Figure 10.** Comparison of the ability to predict responses to grating pattern stimuli between a spiking-based model (grey) and power-based model (white), averaged over all experiments and stimulation amplitudes (mean  $\pm$ SEM) ( $n=40$ ). Asterisks represent a significant difference ( $p < 0.05$ ; Rank sum test). **a)** Coefficient of determination between the model-predicted responses to oriented pattern stimuli and the recorded responses. **b)** Slope of the line of best fit for the correlation described in panel **a**. **c)** Percentage of responses to oriented pattern stimuli that were able to be predicted significantly using bootstrapping analyses.

MUA vs. EP power with anodic-first pulses:  $0.75 \pm 0.04$  vs.  $0.94 \pm 0.12$ ,  $p = 0.08$ . MUA vs. EP power with cathodic-first pulses:  $0.72 \pm 0.05$  vs.  $0.93 \pm 0.10$ ,  $p = 0.03$ ; Rank sum test).

While the percentage of responses that were able to be predicted significantly above chance was generally high for both types of models and all types of stimuli (via bootstrap analysis, Figure 10(c)), the model fitted to MUA was able to predict a significantly higher number of responses for both mixed-phase and anodic-first stimuli compared to the EP power model (MUA vs. EP power with mixed-phase pulses:  $94\% \pm 2.1\%$  vs.  $77\% \pm 5.2\%$ ,  $p = 0.0024$ . MUA vs. EP power with anodic-first pulses:  $85\% \pm 5.4\%$  vs.  $70\% \pm 7.0\%$ ,  $p = 0.129$ . MUA vs. EP power with cathodic-first pulses:  $70\% \pm$

Predicting responses to electrical retinal stimulation

1 5.8% vs.  $77\% \pm 6.1\%$ ,  $p = 0.326$ ; Rank sum test).

2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

2 When predicting responses to single electrode stimulation, coefficients of  
3 determination corresponding to EP power models were significantly higher than those  
4 corresponding to MUA models. However, both were lower than those for multi-electrode  
5 stimulation (MUA vs. EP power:  $r^2 = 0.26 \pm 0.02$  vs.  $0.37 \pm 0.02$ ,  $p = 0.016$ ; Rank sum  
6 test). Despite being low, the majority of both MUA and EP responses to stimuli above  
7 threshold had  $r^2$  values significantly higher than those of the corresponding bootstrapped  
8 distributions (MUA vs. EP power:  $90.6\% \pm 1.20\%$  vs  $92.76\% \pm 0.75\%$ , averaged over  
9 above-threshold stimulus amplitudes and experiments). The slopes of the lines of best fit  
10 for the models fitted to MUA were significantly greater than those fitted to EP power  
11 (MUA vs. EP power:  $0.40 \pm 0.03$  vs.  $0.12 \pm 0.01$ ,  $p < 0.001$ ; Rank sum test). The  
12 percentages of responses able to be predicted accurately by either model were also  
13 relatively low, and not significantly different between the model types (MUA vs. EP  
14 power:  $53\% \pm 3.0\%$  vs.  $45\% \pm 1.5\%$ ,  $p = 0.208$ ; Rank sum test).

Predicting responses to electrical retinal stimulation

## 1 Discussion

2 We have demonstrated that cortical responses to simultaneous stimulation of the retina  
3 are repeatable and can be predicted by a simple linear-nonlinear (LN) model. We have  
4 shown that the model can be optimized using either multi-unit activity (MUA) or power in  
5 the multi-peaked evoked potential (EP), and can be used to predict responses to types of  
6 stimulation that were not used to fit it. The optimized model also provides information  
7 about electrical receptive fields, including the relative effects of each stimulating  
8 electrode on every recording site. These ERFs show that the topographic mapping of  
9 cortical responses to single electrode stimulation (Elfar et al., 2009; Shivdasani et al.,  
10 2012; Wong et al., 2016) is maintained for simultaneous stimulation.

### 11 *Comparison with Previous Studies*

12 Studies have shown that simultaneous multi-electrode stimulation can extend the range  
13 of cortical responses possible from retinal implants beyond what is available with  
14 interleaved single electrode stimulation (Cicione et al., 2012; Matteucci et al., 2013;  
15 Dumm et al., 2014). These studies have characterized the responses of neurons to  
16 particular forms of simultaneous stimulation; however these observations cannot be  
17 generalized to patterns for which the system has not been exposed. Response models  
18 recovered by white noise electrical stimulation go some way towards solving this issue,  
19 since white noise covers a wide range of possible inputs (Chichilnisky, 2001).

20 In terms of visual prostheses, these investigations have been limited to *in vitro*  
21 preparations (Jepson et al., 2014; Maturana et al., 2016). Jepson et al. (2014) showed  
22 that a piecewise linear model could predict the activation of a target cell in the retina,  
23

Predicting responses to electrical retinal stimulation

1  
2  
3 1 however the method was limited to stimulation of small numbers of electrodes at fixed  
4  
5 2 stimulation amplitudes. Maturana et al. (2016) successfully showed that a LN model,  
6  
7 3 mathematically similar to our model, captured the responses of RGCs to simultaneous  
8  
9 4 white noise stimulation of 20 electrodes. While the success of these models aligns with  
10  
11 5 that of our LN model, several important distinctions should be made. In the present  
12  
13 6 study, we have demonstrated successful prediction of activity at multiple concurrent sites  
14  
15 7 in the visual cortex (up to 120 recording channels) in response to stimulation, as  
16  
17 8 opposed to one or two cells at a time (Maturana et al. and Jepson et al. respectively).  
18  
19 9 Additionally, neither study investigated whether the suggested model performed equally  
20  
21 10 well for stimulation that was non-white, as was shown for our model. Also, activation of  
22  
23 11 the visual cortex more closely approximates perception elicited by electrical stimulation  
24  
25 12 of the human retina than activity recorded *in vitro* (von der Heydt and Peterhans, 1989;  
26  
27 13 Gilbert and Wiesel, 1990; Salzman et al., 1990; Knierim et al., 1992; van Wezel et al.,  
28  
29 14 1997), and finally we made use of the same clinical grade electrode array for stimulation  
30  
31 15 as has been trailed in patients (Ayton et al., 2014; Shivdasani et al., 2014).  
32  
33  
34  
35  
36  
37  
38  
39  
40

### 41 *Comparison of MUA-based and EP power-based models*

42  
43  
44 18 Two activity measures in the visual cortex were used to fit separate LN models:  
45  
46 19 MUA and EP power. These activity measures correspond to two different types of  
47  
48 20 cortical processing: MUA reflects the spiking activity of multiple neurons within a radius  
49  
50 21 of 150-300 $\mu$ m from the electrode tip (Gray et al., 1995; Henze et al., 2000); the EP  
51  
52 22 contains both supra-threshold and sub-threshold components, the latter of which  
53  
54 23 extends out over a larger area (Fregnac et al., 1996). The electrically evoked potentials  
55  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

1  
2  
3 1 seen in our study were found to be in a relatively high frequency band compared to  
4  
5 2 traditional local field potentials, and they exhibited a characteristic, positive-going,  
6  
7 3 multi-peaked shape. A number of studies have investigated the origins of these evoked  
8  
9 4 potentials and the factor that influence them (Chang, 1950; Malis and Kruger, 1956;  
10  
11 5 Schoolman and Evarts, 1959; Doty and Grimm, 1962; Burke et al., 1985; Mitzdorf, 1985).  
12  
13 6 Burke et al. (1985) suggested that the first two peaks of this multi-peaked response are  
14  
15 7 due to conduction from Y and X type retinal ganglion cells, respectively (Enroth-Cugell  
16  
17 8 and Robson, 1966; Stone, 1983). Mitzdorf (1985) found that the polarity of the peaks  
18  
19 9 was dependent on the cortical depth of their recording electrode. Thus, the constant  
20  
21 10 polarity of the peaks in our study may be attributed to the fact that all recording  
22  
23 11 electrodes on the penetrating array were the same length (1 mm) and were thus likely to  
24  
25 12 be recording from the same cortical layer. More recently, the EP has been shown to be a  
26  
27 13 viable alternative to MUA when comparing percepts from visual prostheses to those from  
28  
29 14 light stimulation, since a similar EP is present in photic stimulation, albeit with delayed  
30  
31 15 peak latencies due to retinal processing (Nakauchi et al., 2005; Sun et al., 2011). Our  
32  
33 16 findings expand on this, to show that both measures of activity can be the basis of a  
34  
35 17 model for characterizing cortical responses to simultaneous stimulation of the retina.  
36  
37 18 Both model types were able to predict the vast majority of white noise stimuli that were  
38  
39 19 presented in all experiments. In addition, both models displayed high correlation  
40  
41 20 coefficients between recorded and predicted cortical responses to retinal stimuli that  
42  
43 21 were not white, and were able to predict the majority of responses to these stimuli. No  
44  
45 22 consistent differences were found between different receptive field sizes. However,  
46  
47 23 given that we have recorded multi-unit activity as opposed to activity from single  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

1 neurons, it is difficult to make any conclusions regarding the significance of this finding.

2         Some differences were present between the models however; the MUA model  
3 performed significantly better than the EP power model at predicting responses to white  
4 noise, mixed-phase first stimuli and anodic-phase first stimuli. This may be due to a  
5 greater signal to noise ratio for the MUA responses than EP power, which suggests that,  
6 for a model based on EP power, the number of repetitions for each stimulus should be  
7 increased.

8         Neither model performed well in predicting responses to single electrode  
9 stimulation. This may be attributed to the lower thresholds observed when stimulating  
10 across several electrodes simultaneously (Shivdasani et al., 2012), such as during  
11 white-noise stimulation, causing the model to underestimate the current required to  
12 match responses to single electrode stimulation. However, both models could predict  
13 responses to above-threshold single electrode stimulation better than chance,  
14 suggesting that the retinotopic organization is preserved. From a clinical stand-point it is  
15 therefore possible that if the highest safe amplitude of single electrode stimulation is  
16 unable to elicit a percept, simultaneous stimulation of nearby electrodes could elicit a  
17 response in the required area.

18         During the present study, recording channels were only considered for analyses if  
19 they exhibited a monotonic increase in response with increasing stimulation current on  
20 any single retinal electrode. As a result, a significantly greater number of sites were  
21 excluded from the MUA analyses than from EP power analyses. This likely reflects the  
22 requirement that, for recording MUA, the recording site be in close proximity to spiking  
23 cells, whereas estimating EP power does not require direct proximity to cells to register

Predicting responses to electrical retinal stimulation

1  
2  
3 1 an increase. Thus, while the MUA model predicted cortical responses more accurately  
4  
5 2 than the EP power model, the latter had the advantage of characterizing a greater  
6  
7 3 number of recording channels. This suggests that the activity measure on which to train  
8  
9 4 the model could be altered dependent on the requirements of the application, and time  
10  
11 5 available to record responses.  
12  
13

14  
15 6 Clinically, the EP power would be a more attractive cortical measure as currently it  
16  
17 7 is possible to record these EPs with less invasive methods than those required to record  
18  
19 8 MUA; i.e., penetrating electrodes for MUA vs. scalp electrodes. Additionally, EPs have  
20  
21 9 been shown to be more robust for the long term, retaining information even when spikes  
22  
23 10 are lost on the same electrode (Flint et al., 2012). Thus, the EP power is an attractive  
24  
25 11 measure for use with human patients, since it would eliminate the need for a further,  
26  
27 12 invasive procedure, and remain informative in the long term.  
28  
29  
30  
31

32 13

### 33 14 *Considerations for Future Applications*

34  
35  
36 15 The model described in this study is purely spatial in nature owing to the sparse timing  
37  
38 16 used between stimulus pulses. A similar model has been employed previously to model  
39  
40 17 the spatio-temporal dynamics of neurons responding to visual stimulation of the retina  
41  
42 18 (DeAngelis et al., 1993). We expect that by modifying our model to include a temporal  
43  
44 19 dimension in our linear filters, as by Chichilnisky (2001) and Kameneva et al. (2015), the  
45  
46 20 model could be extended to predict temporal dynamics of cortical responses during  
47  
48 21 repetitive, high-rate stimulation, such as perceptual fading. While there is strong  
49  
50 22 evidence suggesting that perceptual fading could be attributed to desensitization  
51  
52 23 observed in retinal ganglion cells (Freeman and Fried, 2011), it could also be attributed  
53  
54  
55  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

1  
2  
3 1 to neural mechanisms within central visual structures. Since in human trials, pulse rates  
4  
5 2 for a suprachoroidal prosthesis have ranged between 50 and 500 pulses per second  
6  
7 3 (pps) (Ayton et al., 2014; Shivdasani et al., 2014) and in other neural systems stimulation  
8  
9 4 rates may be even higher (e.g., stimulation at up to 2000 pps has been studied in  
10  
11 5 cochlear implants (Galvin III and Fu, 2005)), further investigation of the characteristics of  
12  
13 6 adaptation would be of particular interest. However, in pursuit of this goal, one would  
14  
15 7 have to overcome the hurdle of the stimulation artefact present in the recordings. Our  
16  
17 8 stimulus pulses were 1 ms per phase and biphasic, evoking cortical responses lasting  
18  
19 9 approximately 20 ms. The use of stimulus rates higher than 50 pps would not allow  
20  
21 10 analysis of activity with each pulse in the stimulus train without contamination by the  
22  
23 11 stimulus artifact of subsequent stimulus pulses. However, evidence suggests that  
24  
25 12 desensitization of retinal ganglion cells and perceptual fading can also occur at rates  
26  
27 13 lower than 50 pps (Freeman and Fried, 2011; Fornos et al., 2012). At such rates, it would  
28  
29 14 be possible to remove artefacts with the same technique that we have used (Heffer and  
30  
31 15 Fallon, 2008) and analyze the responses between pulses. The model we propose could  
32  
33 16 then be adjusted to incorporate spatiotemporal ERFs (Chichilnisky, 2001).

34  
35  
36  
37  
38  
39  
40  
41 17 Our model also provides an intriguing option for investigation of the effect of  
42  
43 18 photoreceptor degeneration on responses to electrical retinal stimulation. Photoreceptor  
44  
45 19 degeneration in a rat model has been shown to be accompanied by an increased  
46  
47 20 activation threshold to electrical stimulation (Suzuki et al., 2004; O'Hearn et al., 2006) as  
48  
49 21 well as an increase in baseline spiking (Pu et al., 2006; Stasheff, 2008). This may be a  
50  
51 22 contributing factor to the inconsistencies found in the clinical results of simultaneous  
52  
53 23 stimulation. Photoreceptor degeneration also leads to retinal remodeling (Jones et al.,  
54  
55  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

1  
2  
3 1 2003; Marc et al., 2003). Thus, it is likely that the electrical receptive fields of cortical  
4  
5  
6 2 neurons would be different in appearance to those described here.

7  
8 3 Another factor that should be considered in future applications of this method is  
9  
10 4 the effect of anesthesia on cortical activity. Given that anesthesia has been found to  
11  
12 5 reduce the signal-to-noise ratio of visual responses in the cat visual cortex (Livingstone  
13  
14 6 and Hubel, 1981) and preserve the main tuning characteristics of V1 neurons in  
15  
16 7 monkeys (Lamme et al., 1998) and mice (Niell and Stryker, 2010), we expect that the  
17  
18 8 responses to electrical stimulation in an awake cat would be stronger but have similar  
19  
20 9 tuning properties and, therefore, be well suited for application of this method.  
21  
22  
23

24 10 The model we have presented provides novel insights into the effects of  
25  
26 11 interactions between electrodes during simultaneous electrical retinal stimulation. It is  
27  
28 12 expected that with some adjustments, this model could provide a potential approach for  
29  
30 13 choosing optimal stimulation patterns for eliciting specific neural activity (for example,  
31  
32 14 activity more akin to that elicited by visual stimulation). This would entail first fitting the  
33  
34 15 model to responses of the system in question to spatial white noise stimulation. Then, a  
35  
36 16 second optimization algorithm would be used to compute stimulation patterns that would  
37  
38 17 be likely to produce a desired cortical response. In this way, the model could be used to  
39  
40 18 ultimately increase the resolution of the implant.  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50

## 51 20 **Conclusion**

52 21 In this study, we have shown that cortical responses to simultaneous stimulation of the  
53  
54 22 retina are consistent and repeatable. We investigated the effects of electrode  
55  
56 23 interactions of retinal stimulation by creating a linear-nonlinear model of visual cortex  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

1  
2  
3 1 responses. We showed that the model could accurately predict spiking neural responses  
4  
5 2 to both white-noise stimulation and patterned stimulation. The model could also be  
6  
7 3 based on an alternative measure of cortical activity, namely power in the evoked  
8  
9 4 potentials, suggesting that, for clinical applications, a less invasive measure could be  
10  
11 5 applicable. The model we describe provides an effective means of understanding the  
12  
13 6 spatial interactions of retinal stimulation at the level of the cortex, showing that they are  
14  
15 7 predominantly linear and the electrical receptive fields of cortical recording channels are  
16  
17 8 topographically organized. The predictive success of our model shows promise for  
18  
19 9 efficiently determining optimal stimulation paradigms for shaping neural activity with a  
20  
21 10 neural prosthesis.  
22  
23  
24  
25  
26  
27  
28  
29

## 30 **Acknowledgements**

31  
32  
33 13 The authors thank Penny Allen, Jonathan Yeoh, and Chi Luu for surgeries; Carla Abbott,  
34  
35 14 Alice Brandli, Alexia Saunders, Michelle McPhedran, Alison Neil, Dimitra Stathopoulos,  
36  
37 15 Stephanie Epp, and Ceara McGowan for experimental assistance and animal handling;  
38  
39 16 Owen Burns and Vanessa Maxim for manufacturing of the electrodes; and Thomas  
40  
41 17 Spencer, Faith Lamont, Ali Almasi, Felix Aplin, Rosemary Cicione, and Ronald Leung for  
42  
43 18 assistance with data collection. This research was supported by the Australian Research  
44  
45 19 Council through its Special Research Initiative in Bionic Vision Science and Technology  
46  
47 20 awarded to Bionic Vision Australia, the Bertalli Family Foundation to the Bionics Institute,  
48  
49 21 and a project grant from the National Health and Medical Research Council, Australia  
50  
51 22 (GNT#1063093). The Bionics Institute acknowledges the support it receives from the  
52  
53 23 Victorian Government through its Operational Infrastructure Program. KJH was  
54  
55  
56  
57  
58  
59  
60

Predicting responses to electrical retinal stimulation

1 supported by an Australian Postgraduate Award through the Australian Government,  
2 and a postgraduate scholarship from the National Information and Communication  
3 Technology Australia (NICTA - [www.nicta.com.au/](http://www.nicta.com.au/)). NICTA is funded by the Australian  
4 Government as represented by the Department of Broadband, Communications and the  
5 Digital Economy ([www.communications.gov.au/](http://www.communications.gov.au/)) and the Australian Research Council  
6 through the ICT Centre of Excellence program. ANB and YTW acknowledge support  
7 under the Australian Research Council's Discovery Projects funding scheme (Project  
8 DP140104533).

## References

- 11 Auner GW, You R, Siy P, McAllister JP, Talukder M, Abrams GW (2008) Development of a  
12 Wireless High-Frequency Microarray Implant for Retinal Stimulation. *Artificial Sight*  
13 2008:169-186.
- 14 Ayton LN, Blamey PJ, Guymer RH, Luu CD, Nayagam DA, Sinclair NC, Shivdasani MN, Yeoh J,  
15 McCombe MF, Briggs RJ, others (2014) First-in-human trial of a novel suprachoroidal  
16 retinal prosthesis. *PloS one* 9:e115239.
- 17 Barry MP, Dagnelie G, Group AIS (2012) Use of the Argus II retinal prosthesis to improve visual  
18 guidance of fine hand movements. *Investigative Ophthalmology & Visual Science*  
19 53:5095.
- 20 Benjamini Y, Hochberg Y (1995) Controlling the false discovery rate: a practical and powerful  
21 approach to multiple testing. *Journal of the Royal Statistical Society Series B*  
22 (Methodological):289-300.
- 23 Boinagrov D, Pangratz-Fuehrer S, Goetz G, Palanker D (2014) Selectivity of direct and  
24 network-mediated stimulation of the retinal ganglion cells with epi-, sub-and intraretinal  
25 electrodes. *Journal of neural engineering* 11:026008.
- 26 Burke W, Burne J, Martin P (1985) Selective block of Y optic nerve fibres in the cat and the  
27 occurrence of inhibition in the lateral geniculate nucleus. *The Journal of Physiology*  
28 364:81-92.
- 29 Chang H-T (1950) The repetitive discharges of corticothalamic reverberating circuit. *Journal of*  
30 *Neurophysiology*.
- 31 Chichilnisky E (2001) A simple white noise analysis of neuronal light responses. *Network:*  
32 *Computation in Neural Systems* 12:199-213.
- 33 Cicione R, Shivdasani MN, Fallon JB, Luu CD, Allen PJ, Rathbone GD, Shepherd RK, Williams CE  
34 (2012) Visual cortex responses to suprachoroidal electrical stimulation of the retina:  
35 Effects of electrode return configuration. *Journal of Neural Engineering* 9:036009.
- 36 da Cruz L, Coley BF, Dorn J, Merlini F, Filley E, Christopher P, Chen FK, Wuyyuru V, Sahel J,

## Predicting responses to electrical retinal stimulation

- 1  
2  
3 1 Stanga P (2013) The Argus II epiretinal prosthesis system allows letter and word reading  
4 2 and long-term function in patients with profound vision loss. *British Journal of*  
5 3 *Ophthalmology* 0:1-5.  
6 4 DeAngelis GC, Ohzawa I, Freeman R (1993) Spatiotemporal organization of simple-cell receptive  
7 5 fields in the cat's striate cortex. II. Linearity of temporal and spatial summation. *Journal*  
8 6 *of Neurophysiology* 69:1118-1135.  
9 7 Doty RW, Grimm FR (1962) Cortical responses to local electrical stimulation of retina.  
10 8 *Experimental Neurology* 5:319-334.  
11 9 Dumm G, Fallon J, Williams C, Shivdasani M (2014) Virtual electrodes by current steering in  
12 10 retinal prostheses. *Investigative Ophthalmology & Visual Science* 55:8077–8085.  
13 11 Elfar SD, Cottaris NP, Iezzi R, Abrams GW (2009) A cortical (V1) neurophysiological recording  
14 12 model for assessing the efficacy of retinal visual prostheses. *Journal of Neuroscience*  
15 13 *Methods* 180:195-207.  
16 14 Enroth-Cugell C, Robson JG (1966) The contrast sensitivity of retinal ganglion cells of the cat.  
17 15 *The Journal of Physiology* 187:517-552.  
18 16 Flint RD, Lindberg EW, Jordan LR, Miller LE, Slutzky MW (2012) Accurate decoding of reaching  
19 17 movements from field potentials in the absence of spikes. *Journal of neural engineering*  
20 18 9:046006.  
21 19 Fornos AP, Sommerhalder J, da Cruz L, Sahel JA, Mohand-Said S, Hafezi F, Pelizzone M (2012)  
22 20 Temporal Properties of Visual Perception on Electrical Stimulation of the  
23 21 Retina Perception upon Electrical Stimulation of the Retina. *Investigative ophthalmology*  
24 22 *& visual science* 53:2720-2731.  
25 23 Freeman DK, Fried SI (2011) Multiple components of ganglion cell desensitization in response to  
26 24 prosthetic stimulation. *Journal of neural engineering* 8:016008.  
27 25 Fregnac Y, Bringuier V, Chavane F (1996) Synaptic integration fields and associative plasticity of  
28 26 visual cortical cells in vivo. *Journal of Physiology-Paris* 90:367-372.  
29 27 Galvin III JJ, Fu Q-J (2005) Effects of stimulation rate, mode and level on modulation detection  
30 28 by cochlear implant users. *Journal of the Association for Research in Otolaryngology*  
31 29 6:269-279.  
32 30 Gilbert CD, Wiesel TN (1990) The influence of contextual stimuli on the orientation selectivity of  
33 31 cells in primary visual cortex of the cat. *Vision Research* 30:1689-1701.  
34 32 Gray CM, Maldonado PE, Wilson M, McNaughton B (1995) Tetrodes markedly improve the  
35 33 reliability and yield of multiple single-unit isolation from multi-unit recordings in cat  
36 34 striate cortex. *Journal of Neuroscience Methods* 63:43-54.  
37 35 Habib AG, Cameron MA, Suaning GJ, Lovell NH, Morley JW (2013) Spatially restricted electrical  
38 36 activation of retinal ganglion cells in the rabbit retina by hexapolar electrode return  
39 37 configuration. *Journal of Neural Engineering* 10:036013.  
40 38 Heffer LF, Fallon JB (2008) A novel stimulus artifact removal technique for high-rate electrical  
41 39 stimulation. *Journal of Neuroscience Methods* 170:277-284.  
42 40 Henze DA, Borhegyi Z, Csicsvari J, Mamiya A, Harris KD, Buzsáki G (2000) Intracellular features  
43 41 predicted by extracellular recordings in the hippocampus in vivo. *Journal of*  
44 42 *Neurophysiology* 84:390-400.  
45 43 Horsager A, Greenberg RJ, Fine I (2010) Spatiotemporal interactions in retinal prosthesis  
46 44 subjects. *Investigative Ophthalmology & Visual Science* 51:1223-1233.

## Predicting responses to electrical retinal stimulation

- 1  
2  
3 1 Humayun MS, de Juan Jr E, Weiland JD, Dagnelie G, Katona S, Greenberg RJ, Suzuki S (1999)  
4 2 Pattern electrical stimulation of the human retina. *Vision Research* 39:2569-2576.  
5 3 Jensen RJ, Rizzo JF (2006) Thresholds for activation of rabbit retinal ganglion cells with a  
6 4 subretinal electrode. *Experimental eye research* 83:367-373.  
7 5 Jepson L, Hottowy P, Gunning D, Mathieson K, Dabrowski W, Litke A, Chichilnisky E (2014)  
8 6 Spatially Patterned Electrical Stimulation to Enhance Resolution of Retinal Prostheses.  
9 7 *The Journal of Neuroscience* 34:4871-4881.  
10 8 Jones BW, Watt CB, Frederick JM, Baehr W, Chen CK, Levine EM, Milam AH, Lavail MM, Marc RE  
11 9 (2003) Retinal remodeling triggered by photoreceptor degenerations. *Journal of*  
12 10 *Comparative Neurology* 464:1-16.  
13 11 Kameneva T, Abramian M, Zarelli D, Nęsić D, Burkitt AN, Meffin H, Grayden DB (2015) Spike  
14 12 history neural response model. *Journal of computational neuroscience* 38:463-481.  
15 13 Knierim JJ, Van Essen DC, others (1992) Neuronal responses to static texture patterns in area V1  
16 14 of the alert macaque monkey. *Journal of Neurophysiology* 67:961-980.  
17 15 Kotecha A, Zhong J, Stewart D, da Cruz L (2014) The Argus II prosthesis facilitates reaching and  
18 16 grasping tasks: a case series. *BMC Ophthalmology* 14:71.  
19 17 Lamme VA, Zipser K, Spekreijse H (1998) Figure-ground activity in primary visual cortex is  
20 18 suppressed by anesthesia. *Proceedings of the National Academy of Sciences*  
21 19 95:3263-3268.  
22 20 Livingstone MS, Hubel DH (1981) Effects of sleep and arousal on the processing of visual  
23 21 information in the cat. *Nature* 291:554-561.  
24 22 Malis LI, Kruger L (1956) Multiple response and excitability of cat's visual cortex. *Journal of*  
25 23 *Neurophysiology* 19:172-186.  
26 24 Marc RE, Jones BW, Watt CB, Strettoi E (2003) Neural Remodeling in Retinal Degeneration.  
27 25 *Progress in Retinal and Eye Research* 22:607-655.  
28 26 Matteucci PB, Chen SC, Tsai D, Dodds CWD, Dokos S, Morley JW, Lovell NH, Suaning GJ (2013)  
29 27 Current Steering in Retinal Stimulation via a Quasimonopolar Stimulation Paradigm.  
30 28 *Investigative Ophthalmology & Visual Science* 54:4307-4320.  
31 29 Maturana MI, Apollo NV, Hadjinicolaou AE, Garrett DJ, Cloherty SL, Kameneva T, Grayden DB,  
32 30 Ibbotson MR, Meffin H (2016) A Simple and Accurate Model to Predict Responses to  
33 31 Multi-electrode Stimulation in the Retina. *PLOS Comput Biol* 12:e1004849.  
34 32 Mitra PP, Pesaran B (1999) Analysis of dynamic brain imaging data. *Biophysical Journal*  
35 33 76:691-708.  
36 34 Mitzdorf U (1985) Current source-density method and application in cat cerebral cortex:  
37 35 investigation of evoked potentials and EEG phenomena. *Physiological Reviews*  
38 36 65:37-100.  
39 37 Nakauchi K, Fujikado T, Kanda H, Morimoto T, Choi JS, Ikuno Y, Sakaguchi H, Kamei M, Ohji M,  
40 38 Yagi T (2005) Transretinal electrical stimulation by an intrascleral multichannel electrode  
41 39 array in rabbit eyes. *Graefes Archive for Clinical and Experimental Ophthalmology*  
42 40 243:169-174.  
43 41 Niell CM, Stryker MP (2010) Modulation of visual responses by behavioral state in mouse visual  
44 42 cortex. *Neuron* 65:472-479.  
45 43 O'Hearn TM, Sadda SR, Weiland JD, Maia M, Margalit E, Humayun MS (2006) Electrical  
46 44 stimulation in normal and retinal degeneration (rd1) isolated mouse retina. *Vision*

## Predicting responses to electrical retinal stimulation

- 1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60
- 1 Research 46:3198-3204.
- 2 Pillow JW, Paninski L, Uzzell VJ, Simoncelli EP, Chichilnisky E (2005) Prediction and decoding of  
3 retinal ganglion cell responses with a probabilistic spiking model. *The Journal of*  
4 *Neuroscience* 25:11003-11013.
- 5 Pu M, Xu L, Zhang H (2006) Visual response properties of retinal ganglion cells in the royal  
6 college of surgeons dystrophic rat. *Investigative Ophthalmology & Visual Science*  
7 47:3579-3585.
- 8 Rizzo JF, Wyatt J, Loewenstein J, Kelly S, Shire D (2003) Perceptual Efficacy of Electrical  
9 Stimulation of Human Retina with a Microelectrode Array during Short-Term Surgical  
10 Trials. *Investigative Ophthalmology & Visual Science* 44:5362-5369.
- 11 Salzman CD, Britten KH, Newsome WT (1990) Cortical microstimulation influences perceptual  
12 judgements of motion direction. *Nature* 346:174-177.
- 13 Schoolman A, Everts EV (1959) Responses to lateral geniculate radiation stimulation in cats with  
14 implanted electrodes. *Journal of Neurophysiology* 22:112-129.
- 15 Schwartz G, Rieke F (2011) Nonlinear spatial encoding by retinal ganglion cells: when  $1 + 1 \neq$   
16 2. *The Journal of General Physiology* 138:283-290.
- 17 Shivdasani MN, Fallon JB, Luu CD, Cicione R, Allen PJ, Morley JW, Williams CE (2012) Visual  
18 Cortex Responses to Single-and Simultaneous Multiple-Electrode Stimulation of the  
19 Retina: Implications for Retinal Prostheses. *Investigative Ophthalmology & Visual*  
20 *Science* 53:6291-6300.
- 21 Shivdasani MN, Sinclair NC, Dimitrov PN, Varsamidis M, Ayton LN, Luu CD, Perera T, McDermott  
22 HJ, Blamey PJ (2014) Factors Affecting Perceptual Thresholds in a Suprachoroidal Retinal  
23 Prosthesis Factors Affecting Retinal Prosthesis Thresholds. *Investigative ophthalmology*  
24 *& visual science* 55:6467-6481.
- 25 Stasheff SF (2008) Emergence of sustained spontaneous hyperactivity and temporary  
26 preservation of OFF responses in ganglion cells of the retinal degeneration (rd1) mouse.  
27 *Journal of Neurophysiology* 99:1408-1421.
- 28 Stingl K, Bartz-Schmidt KU, Besch D, Braun A, Bruckmann A, Gekeler F, Greppmaier U, Hipp S,  
29 Hörtdörfer G, Kernstock C (2013) Artificial vision with wirelessly powered subretinal  
30 electronic implant alpha-IMS. *Proceedings of the Royal Society of London B: Biological*  
31 *Sciences* 280:20130077.
- 32 Stingl K, Bartz-Schmidt KU, Besch D, Chee CK, Cottrill CL, Gekeler F, Groppe M, Jackson TL,  
33 MacLaren RE, Koitschev A (2015) Subretinal Visual Implant Alpha IMS—Clinical trial  
34 interim report. *Vision research* 111:149-160.
- 35 Stone J (1983) On the Understanding of Visual Processing in the Diencephalon. In: *Parallel*  
36 *Processing in the Visual System*, pp 149-193: Springer.
- 37 Sun J, Lu Y, Cao P, Li X, Cai C, Chai X, Ren Q, Li L (2011) Spatiotemporal properties of  
38 multip peaked electrically evoked potentials elicited by penetrative optic nerve  
39 stimulation in rabbits. *Investigative Ophthalmology & Visual Science* 52:146-154.
- 40 Suzuki S, Humayun MS, Weiland JD, Chen S-J, Margalit E, Piyathaisere DV, de Juan JE (2004)  
41 Comparison of electrical stimulation thresholds in normal and retinal degenerated  
42 mouse retina. *Japanese Journal of Ophthalmology* 48:345-349.
- 43 Thomson DJ (1982) Spectrum estimation and harmonic analysis. *Proceedings of the IEEE*  
44 70:1055-1096.

## Predicting responses to electrical retinal stimulation

- 1  
2  
3 1 Tusa R, Palmer L, Rosenquist A (1978) The retinotopic organization of area 17 (striate cortex) in  
4 2 the cat. *Journal of Comparative Neurology* 177:213-235.  
5 3  
6 3 van Wezel RJ, Lankheet MJ, Fredericksen R, Verstraten FA, van de Grind W (1997) Responses of  
7 4 complex cells in cat area 17 to apparent motion of random pixel arrays. *Vision Research*  
8 5 37:839-852.  
9 6  
10 6 Villalobos J, Allen PJ, McCombe MF, Ulaganathan M, Zamir E, Ng DC, Shepherd RK, Williams CE  
11 7 (2012) Development of a surgical approach for a wide-view suprachoroidal retinal  
12 8 prosthesis: evaluation of implantation trauma. *Graefe's Archive for Clinical and*  
13 9 *Experimental Ophthalmology* 250:399-407.  
14 10  
15 10 von der Heydt R, Peterhans E (1989) Mechanisms of contour perception in monkey visual  
16 11 cortex. I. Lines of pattern discontinuity. *The Journal of Neuroscience* 9:1731-1748.  
17 12  
18 12 Wilke R, Gabel V-P, Sachs H, Schmidt K-UB, Gekeler F, Besch D, Szurman P, Stett A, Wilhelm B,  
19 13 Peters T (2011) Spatial resolution and perception of patterns mediated by a subretinal  
20 14 16-electrode array in patients blinded by hereditary retinal dystrophies. *Investigative*  
21 15 *Ophthalmology & Visual Science* 52:5995-6003.  
22 16  
23 16 Wong YT, Halupka K, Kameneva T, Cloherty SL, Grayden DB, Burkitt AN, Meffin H, Shivdasani  
24 17 MN (2016) Spectral distribution of local field potential responses to electrical  
25 18 stimulation of the retina. *Journal of neural engineering* 13:036003.  
26 19  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60